

**DESCRIPTION**

**Source** Mouse myeloma cell line, NS0-derived  
Ala121-Gln730, with a C-terminal 6-His tag  
Accession # NP\_001190

**N-terminal Sequence Analysis** Ala121

**Predicted Molecular Mass** 70.5 kDa

**SPECIFICATIONS**

**SDS-PAGE** 84 kDa, reducing conditions

**Activity** Measured by its ability to cleave a fluorogenic peptide substrate, Mca-YVADAPK(Dnp)-OH (Catalog # ES007).  
The specific activity is >4 pmol/min/μg, as measured under the described conditions. See Activity Assay Protocol on www.RnDSystems.com

**Endotoxin Level** <1.0 EU per 1 μg of the protein by the LAL method.

**Purity** >95%, by SDS-PAGE under reducing conditions and visualized by silver stain.

**Formulation** Lyophilized from a 0.2 μm filtered solution in HEPES, Sodium Sulfate and ZnCl<sub>2</sub>. See Certificate of Analysis for details.

**Activity Assay Protocol**

- Materials**
- Assay Buffer: 25 mM HEPES, 0.1% Brij-35 (w/v), pH 7.5
  - Recombinant Human BMP-1/PCP (rhBMP-1) (Catalog # 1927-ZN)
  - Fluorogenic Peptide Substrate: MCA-Tyr-Val-Ala-Asp-Ala-Pro-Lys(DNP)-OH (Catalog # ES007)
  - F16 Black Maxisorp Plate (Nunc, Catalog # 475515)
  - Fluorescent Plate Reader (Model: SpectraMax Gemini EM by Molecular Devices) or equivalent

- Assay**
1. Dilute rhBMP-1 to 20 ng/μL in Assay Buffer.
  2. Dilute substrate to 20 μM in Assay Buffer.
  3. Load into a black well plate 50 μL of 20 ng/μL rhBMP-1 and start the reaction by adding 50 μL of 20 μM Substrate. Include a Substrate Blank containing Assay Buffer in place of rhBMP-1.
  4. Read at excitation and emission wavelengths of 320 nm and 405 nm (top read), respectively, in kinetic mode for 5 minutes.
  5. Calculate specific activity:

$$\text{Specific Activity (pmol/min/}\mu\text{g)} = \frac{\text{Adjusted } V_{\text{max}}^* \text{ (RFU/min)} \times \text{Conversion Factor}^{**} \text{ (pmol/RFU)}}{\text{amount of enzyme (}\mu\text{g)}}$$

\*Adjusted for Substrate Blank

\*\*Derived using calibration standard MCA-Pro-Leu-OH (Bachem, Catalog # M-1975).

- Final Assay Conditions** Per Well:
- rhBMP-1: 1 μg
  - Substrate: 10 μM

**PREPARATION AND STORAGE**

**Reconstitution** Reconstitute at 100 μg/mL in sterile 25 mM HEPES and 0.01% (w/v) Brij-35, pH 7.5.

**Shipping** The product is shipped at ambient temperature. Upon receipt, store it immediately at the temperature recommended below.

- Stability & Storage** Use a manual defrost freezer and avoid repeated freeze-thaw cycles.
- 6 months from date of receipt, -20 to -70 °C as supplied.
  - 3 months, -20 to -70 °C under sterile conditions after reconstitution.

**BACKGROUND**

Bone morphogenetic protein 1 (BMP-1), also known as procollagen C-proteinase (PCP), is a zinc protease of the astacin family (1, 2). BMP-1/PCP plays a key role in formation of extracellular matrix (ECM) by converting precursor proteins into their mature and functional forms. The precursor proteins identified as substrates for BMP-1/PCP include collagens, biglycan, laminin 5, dentin matrix protein-1, and lysyl oxidase (3). There are six alternatively spliced forms known to be derived from the BMP-1 gene, and isoform 1 consisting of residues 1 to 730 was expressed. The secreted and purified protein does not contain the signal peptide (amino acid residues 1-22) and pro domain (residues 23-120), but contain protease (residues 121-321), CUB I (residues 322-434), CUB II (residues 435-546), EGF-like (residues 547-588) and CUB III (residues 591-703) domains. The pro domain is apparently cleaved by a furin-like proprotein convertase (4). The purified BMP-1/PCP is an active protease and its peptidase activity can be determined as described above. The purified BMP-1/PCP is predicted to possess procollagen C-proteinase activity because it contains the minimal domain structure required (5).

**References:**

1. Wozney, J.M. *et al.* (1988) *Science* **242**:1528.
2. Bond, J.S. and R.J. Beynon (1995) *Protein Sci.* **4**:1247.
3. Steiglitz, B.M. *et al.* (2004) *J. Biol. Chem.* **279**:980.
4. Leighton, M. and K.E. Kadler (2003) *J. Biol. Chem.* **278**:18478.
5. Hartigan, N. *et al.* (2003) *J. Biol. Chem.* **278**:18045.